## Quick fish sampling guide for disease diagnostics

## Histology sampling guide

Step 1: Sample collection


For fry or eggs $<1 \mathrm{~cm}$ :
For swim-up fry or fertilized eggs $<1 \mathrm{~cm}$ : Place 3-5 whole fry and/or eggs in 10\% neutral buffered formalin (NBF) fixative at a ratio of fish to fixative of 1:10.


Avoid the use of tubes with narrow bottoms, especially conical, as tissues do not mix adequately with fixative. Better to use flat bottom tubes.For fingerlings $1-5 \mathrm{~cm}$ : Cut off gill opercula. Open abdomen. Pull out viscera to protrude slightly from opening to expose internal organs. Place fish in 10\% NBF fixative at a ratio of fish to fixative of 1:10. Caution: Do not to cut the intestines.


15 ml

For fish > 5 cm :
Dissect a $5 \times 5 \mathrm{~mm}$ section (clean cuts) for each organ (take whole organ if $<5 \mathrm{~mm}$ thick). For standard histology, collect from brain, gill, eye, heart, pyloric caeca, stomach, intestine, spleen, liver, kidney and a representative lesion if any (see below for more details).


Place organ specimens in 10\% NBF fixative at a ratio of fish to fixative of 1:10.



Step 2: Fixation
Immerse $5 \times 5 \mathrm{~mm}$ tissue sections in 10\% NBF fixative at a 1:10 ratio.
Store at room temperature away from sun for 24-48 hours.


