



QUARTERLY AQUATIC ANIMAL DISEASE REPORT (Asia and Pacific Region)

July – September 2013

Published by the

Network of Aquaculture Centres in Asia-Pacific

Suraswadi Building, Department of Fisheries Kasetsart University Campus, Ladyao, Jatujak Bangkok 10900, Thailand Food and Agriculture Organization of the United Nations

> Viale delle Terme di Caracalla Rome 00100 Italy

Quarterly Aquatic Animal Disease Report (Asia-Pacific Region) - 2013/3

Network of Aquaculture Centres in Asia-Pacific and Food and Agriculture Organization of the United Nations. Oct. 2014. *Quarterly Aquatic Animal Disease Report (Asia and Pacific Region)*, 2013/3, July – August 2013. NACA: Bangkok, Thailand.

Contents

Foreword	iv
Reports Received by the NACA Secretariat	
Australia	1
Hong Kong	5
India	7
Indonesia	
2nd Quarter	9
3rd Quarter	12
Iran	16
Japan	18
Korea, Republic of	20
Lao PDR	22
Malaysia	24
Myanmar	28
Nepal	30
Philippines	32
Singapore	35
Sri Lanka	38
Vietnam	
2nd Quarter	41
3rd Quarter	44
List of Diseases under the Asia-Pacific Quarterly Aquatic Animal Disease Report	47
Recent related publications	48
List of National Coordinators	51
Instructions on how to fill in the Quarterly Aquatic Animal Disease Report	55

Foreword

OIE Tool for the Evaluation of Performance of Aquatic Veterinary Services

Paris - In this era of globalisation, the development and growth of many countries, as well as the prevention and control of major biological disasters, depends on the performance of their agricultural and food policies and economies, and this, in turn, directly relates to the quality of their Veterinary Services (VS).

Strengthening of VS to help them comply with OIE international standards for quality and evaluation requires active participation and investment by both the public and the private sector. The World Organisation for Animal Health (OIE) has refined an Evaluation Tool which is designed to assist VS to establish their current level of performance, to identify gaps and weaknesses in their ability to comply with OIE international standards, to form a shared vision with stakeholders (including the private sector) and to establish priorities and carry out strategic initiatives.

The production of and trade in aquatic animals and their products is of increasing importance and the aquaculture sector is growing fast in response to the strong and growing global demand for high quality protein. In some countries the VS are the competent authority for aquatic animal health services (AAHS), but in some countries other agencies of government hold this responsibility.

Regardless of whether veterinarians are involved in the AAHS, it is clear that the general principles for quality would be similar to those that apply to VS. For example, appropriate legislation and good governance are required to support AAHS in complying with OIE requirements, including for disease detection, reporting and control.

The application of the PVS Tool to the evaluation of AAHS commenced in 2009 when the OIE undertook a pilot mission in Vietnam. Following this mission and several subsequent missions, it was clear that a stand-alone tool for the evaluation of an AAHS was necessary due to differences between the terrestrial and aquatic sectors. The OIE has developed the OIE Tool for the Evaluation of Performance of Veterinary Services and/or Aquatic Animal Health Services ("OIE PVS Tool: Aquatic") which is based on the sixth edition of the OIE PVS Tool and includes amendments to some critical competencies (CC) and Levels of Advancement so that the Tool is more appropriate for the evaluation of the performance of AAHS.

Applying the "OIE PVS Tool: Aquatic"

To establish the current level of performance, form a shared vision, establish priorities and carry out strategic initiatives, six to thirteen critical competencies have been elaborated for each of the four fundamental components. For each critical competency, qualitative levels of advancement are described. A higher level of advancement assumes that the VS / AAHS are complying with the preceding (non 1) levels (i.e. level 3 assumes compliance with level 2 criteria; level 5

assumes compliance with level 4 and preceding criteria; etc.). Additional critical competencies might be added with the evolution of these tools.

The OIE has provided a Manual for Assessors, containing information and procedures relevant to the conduct of an OIE PVS Evaluation, including a list of suggested indicators used by PVS assessors. Chapter 3.1. of the Aquatic Animal Health Code (Aquatic Code) provides a legal base for the OIE quality requirements for AAHS where these are not covered by the Veterinary Services.

The benefits and outcomes of using the OIE PVS Tool and the OIE PVS Tool: Aquatic include:

• An indication of overall performance for each of the four components and a relative performance rating within each of the critical competencies;

• A basis for comparing the performance of the VS / AAHS with that of other relevant government services in the region or globally, in order to explore areas for cooperation or negotiation;

• Providing the basis for carrying out a process of verifying compliance with the OIE standards and assessments of VS / AAHS by external or independent bodies under the guidelines and auspices of the OIE;

• Where gaps in the legislative framework are identified in the course of a PVS Evaluation, through the conduct of an OIE Veterinary Legislation Identification Mission, obtaining an indication of the specific actions needed to modernise the veterinary legislation in compliance with OIE standards. Where relevant, the next step is a memorandum of undertaking between the OIE and the Member of specific activities on legislation;

• Through the conduct of a specific follow up, i.e. the OIE PVS Gap Analysis, helping countries to identify priorities and present justifications when applying for national and/or international financial support (loans and/or grants) from national governments or international donors;

• Providing a basis for establishing a routine monitoring and follow up mechanism on the overall level of performance of the VS / AAHS over time;

• Helping to determine the benefits and costs of investing in VS / AAHS and, through the conduct of specific follow up activities, identifying the actions and securing the investments that are needed to help improve compliance with the OIE standards for Good Governance.

See <u>http://www.oie.int/en/support-to-oie-members/pvs-evaluations/oie-pvs-tool/</u> for more information and to download a copy of the "OIE PVS Tool: Aquatic."

Reports Received by the NACA Secretariat

Country: AUSTRALIA

Item Disease status ^{a/}			<u>ı/</u>	Level of	Epidemiological
DISEASES PREVALENT IN THE REGION		Month		Level of diagnosis	comment
FINFISH DISEASES	July	August	September		numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	-(2012)	-(2012)	-(2012)		1
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	-(2012)	+	-(2013)	III	2
6. Red seabream iridoviral disease	0000	0000	0000		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000		
9. Viral encephalopathy and retinopathy	-(2013)	-(2013)	-(2013)		3
10.Enteric septicaemia of catfish	(2011)	(2011)	(2011)		4
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	-(2013)	-(2013)	-(2013)		5
3. Infection with abalone herpes-like virus	-(2011)	-(2011)	-(2011)		6
4. Infection with <i>Xenohaliotis californiensis</i>	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with <i>Marteilioides chungmuensis</i>	0000	0000	0000		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	0000	0000	0000		
3. Yellowhead disease	0000	0000	0000		
4. Infectious hypodermal and haematopoietic necrosis	-(2008)	-(2008)	-(2008)		7
5. Infectious myonecrosis	0000	0000	0000		
6. White tail disease (MrNV)	-(2008)	-(2008)	-(2008)		8
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		
9. Monodon slow growth syndrome	0000	0000	0000		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	***	***	***		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	-(2008)	-(2008)	-(2008)		9
2. Infection with <i>Batrachochytrium dendrobatidis</i>	-(2013)	-(2013)	-(2013)		10
ANY OTHER DISEASES OF IMPORTANCE	< - /		()		
1. Virus isolated from Atlantic salmon	-(2013)	-(2013)	+	III	11
2.	~ /	× /			

nfish:	D BY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus mar	inus.	
	eans: Crayfish plague (Aphanomyces astaci).		
	Channel catfish virus disease		
Pleas	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	***	No information available
	Serological evidence and/or isolation of causative agent but	0000	Never reported
+?	no clinical diseases	-	Not reported (but disease is known to occur)
+?	no chincaí diseases		

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	Epizootic haematopoietic necrosis was not reported this period despite passive surveillance in Victoria (last reported 2012), the Australian Capital Territory (last reported 2011), New South Wales (last reported 2009) and South Australia (last reported 1992). Passive surveillance and never reported in the Northern Territory, Queensland, Tasmania and Western Australia.
	Epizootic ulcerative syndrome 1. Reported in Queensland in August, screening;
	 Species affected – Spikey bream (<i>Acanthopagrus sp.</i>); Clinical signs – skin ulceration on the flank of several spikey bream caught Pathogen – Aphanomyces invadans; Mortality rate – Nil,
2	 Economic loss – Nil; Geographic extent – Limited to waters close to Jumpinpin in southeast Queensland; Containment measures – Not applicable,; Laboratory confirmation – Histopathology; Publications – None.
	Epizootic ulcerative syndrome is known to have occurred previously in New South Wales (last reported 2012), the Northern Territory (last reported 2012), Victoria (last reported 2012), Western Australia (last reported 2012) and South Australia (last reported 2008). Passive surveillance and never reported in Tasmania. No information available in the Australian Capital Territory.
3	Viral encephalopathy and retinopathy was not reported this period despite passive surveillance in the Northern Territory (last reported 2013), Western Australia (last reported 2013), Queensland (last reported 2013), New South Wales (last reported 2010), South Australia (last reported 2010) and Tasmania (last reported 2000). Passive surveillance and never reported in Victoria. No information available in the Australian Capital Territory.

4	Enteric septicaemia of catfish was not reported this period and has never been reported from wild fish in Australia. Passive surveillance and reported previously in the Northern Territory [in a closed aquarium facility also holding imported ornamental fish] (last reported 2011), Queensland (last reported 2008) and Tasmania (last reported 2001) in zebrafish (<i>Brachydanio rerio</i>) held in PC2 containment facilities. Passive surveillance and never reported in New South Wales, South Australia, Victoria or Western Australia. No information available this period in the Australian Capital Territory.
5	Infection with <i>Perkinsus olseni</i> was not reported this quarter despite passive surveillance in South Australia (last reported 2013), New South Wales (last reported 2005). Not reported despite targeted surveillance in Western Australia (last reported 2003). Passive surveillance and never reported in the Northern Territory, Queensland, Tasmania and Victoria. No information available for the Australian Capital Territory (suceptible species not present and no marine water responsibility).
6	Infection with abalone herpesvirus (abalone viral ganglioneuritis) was not reported this period despite targeted surveillance in Tasmania (last reported 2011) and passive surveillance in New South Wales (last reported 2011 and eradicated following detection in contained commercial live-holding facilities), and Victoria (last reported 2010). Passive surveillance and never reported in the Northern Territory, Queensland, South Australia and Western Australia. No information available this period for the Australian Capital Territory (no marine water responsibility).
7	Infectious hypodermal and haematopoietic necrosis virus was not reported this period despite passive surveillance in Queensland (last reported 2008) and Northern Territory (last reported 2003). Passive surveillance and never reported in New South Wales, South Australia, Victoria and Western Australia. No information available this period in the Australian Capital Territory (no marine responsibility) and Tasmania (susceptible species not present).
8	White tail disease was not reported this period despite passive surveillance in Queensland (last reported 2008). Passive surveillance and never reported from the Australian Capital Territory, New South Wales, the Northern Territory, South Australia, Victoria and Western Australia. No information available this period in Tasmania.
9	Infection with ranavirus was not reported this period despite passive surveillance in the Northern Territory (last reported 2008, prior to official reporting for ranavirus). Suspected but not confirmed through passive surveillance in Queensland. Passive surveillance and never reported in Tasmania. No information available this period in the Australian Capital Territory, New South Wales, South Australia, Victoria and Western Australia.
10	Infection with <i>Batrachochytrium dendrobatidis</i> was not reported this period despite targeted surveillance in Tasmania (last reported 2013), passive surveillance in Victoria (last reported 2011) and Western Australia (last reported 2008). Suspected but not confirmed through passive surveillance in Queensland. No information available this period in the Australian Capital Territory, New South Wales, the Northern Territory, and South Australia.
11	Virus isolated from Atlantic salmon. A novel virus has been detected in association with mortalities of cage- cultured Atlantic salmon in south eastern Tasmania. The virus was initially considered to be an incidental finding; however, some sporadic detections have been made in association with variable mortality of smolt. Viral ultrastructure resembles that of the Orthomyxoviridae. Further characterisation, including sequencing is pending. Infectious salmon anemia virus has been excluded through OIE recommended assays. Investigations are on-going to characterise the virus, better understand its epidemiology, and determine its significance as a pathogen of salmon.

Quarterly Aquatic Animal Disease Report (Asia-Pacific Region) - 2013/3

Country: HONG KONG SAR

Item		Disease status ^a	/		<u> </u>
	SEASES PREVALENT IN THE REGION Month			Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases	j j				
1. Epizootic haematopoietic necrosis	0000	0000	0000	II	
2. Infectious haematopoietic necrosis	0000	0000	0000	III	
3. Spring viraemia of carp	0000	0000	0000	III	
4. Viral haemorrhagic septicaemia	0000	0000	0000	III	
5. Epizootic ulcerative syndrome	0000	0000	0000	III	
6. Red seabream iridoviral disease	-	+	-	III	1,2
7. Koi herpesvirus disease	-	-	-	III	1
Non OIE-listed diseases					
8. Grouper iridoviral disease	-	-	-	III	
9. Viral encephalopathy and retinopathy	-	-	-	III	
10.Enteric septicaemia of catfish	0000	0000	0000	II	
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000	II	
2. Infection with Perkinsus olseni	0000	0000	0000	II	
3. Infection with abalone herpes-like virus	0000	0000	0000	II	
4. Infection with Xenohaliotis californiensis					
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000	II	
6. Acute viral necrosis (in scallops)	0000	0000	0000	II	
7. Akoya oyster disease	0000	0000	0000	II	
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000	III	
2. White spot disease	-	-	-	III	
3. Yellowhead disease	0000	0000	0000	III	
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000	II	
5. Infectious myonecrosis	0000	0000	0000	II	
6. White tail disease (MrNV)	0000	0000	0000	II	
7. Necrotising hepatopancreatitis	0000	0000	0000	II	
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (Panulirus spp.)	0000	0000	0000	II	
9. Monodon slow growth syndrome	0000	0000	0000	II	
10. Acute hepatopancreatic necrosis syndrome (AHPNS)					
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000	II	
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000	II	
ANY OTHER DISEASES OF IMPORTANCE					
1.					

ISTED	SES PRESUMED EXOTIC TO THE REGION ^b D BY THE OIE		
	Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . s: Infection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i>	inus	
	eans: Crayfish plague (Aphanomyces astaci).	mus.	
OT LI	ISTED BY THE OIE		
infish:	Channel catfish virus disease		
Pleas	e use the following symbols:	~	
+	Disease reported or known to be present	+() ***	Occurrence limited to certain zones
+?	Serological evidence and/or isolation of causative agent but	0000	No information available
1.	no clinical diseases	0000	Never reported Not reported (but disease is known to occur)
	Suspected by reporting officer but presence not confirmed	(year)	Year of last occurrence
	Suspected by reporting officer but presence not confirmed	(year)	1

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	Infectious spleen and kidney necrosis virus was detected in a group of jade perches with 2% mortality and 2% morbidity reported.
2	Infectious spleen and kidney necrosis virus and red seabream iridovirus were detected in a group of giant groupers with 20% mortality and 50% morbidity reported.
3	

Country: INDIA

T4		Disease status ²		<u> </u>	
Item	EASES PREVALENT IN THE REGION Month			Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases	j - c				
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	-	-	-		
6. Red seabream iridoviral disease	0000	0000	0000		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000		
9. Viral encephalopathy and retinopathy	0000	0000	0000		
10.Enteric septicaemia of catfish	0000	0000	0000		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	-	-	-		
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	+()	+()	+()	Ι	1,2
3. Yellowhead disease	***	***	***		
4. Infectious hypodermal and haematopoietic necrosis	***	***	***		
5. Infectious myonecrosis	0000	0000	0000		
6.White tail disease (MrNV)	-	-	-		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		
9. Monodon slow growth syndrome	-	-	-		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

ctious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . fection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> : Crayfish plague (<i>Aphanomyces astaci</i>).	inus.	
: Cravfish plague (Aphanomyces astaci).		
the following symbols:		
	+()	Occurrence limited to certain zones
Disease reported or known to be present	***	No information available
Serological evidence and/or isolation of causative agent but	0000	Never reported
no clinical diseases	-	Not reported (but disease is known to occur)
Suspected by reporting officer but presence not confirmed	(vear)	Year of last occurrence
, ,	Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases	nnel catfish virus disease the following symbols: Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	White spot disease (WSD): Reported from very limited areas in Bhimavaram and Vijayiwada districts of Andhra Pradesh, Udupi and Uttar Kannada districts of Karnataka, Thane district of Maharashta, and Nagapattinam district of Tamil Nadu during different months under the reporting period.
2	P. monodon and L. vannamei were affected by WSD
3	

Country: INDONESIA

Period: <u>April - June 2013</u>

Item Disease status $\frac{a}{a}$					Epidemiological
DISEASES PREVALENT IN THE REGION				Level of	comment
FINFISH DISEASES	April	May	June	diagnosis	numbers
OIE-listed diseases	¹				
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	0000	0000	0000		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	+()	***	+()	III	1
Non OIE-listed diseases					
8. Grouper iridoviral disease	+()	***	***	II	2
9. Viral encephalopathy and retinopathy	+()	***	***	III	3
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	0000	0000	0000		
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	***	***	***		
2. White spot disease	+()	***	***	III	4
3. Yellowhead disease	***	***	***		
4. Infectious hypodermal and haematopoietic necrosis	***	***	***		
5. Infectious myonecrosis	+()	***	***	III	5
6. White tail disease (MrNV)	0000	0000	0000		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		
9. Monodon slow growth syndrome	0000	0000	0000		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

nfish:	DBY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus man	rinus.	
	eans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE		
	Channel catfish virus disease		
Please	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	***	No information available
+?	Serological evidence and/or isolation of causative agent but	0000	Never reported
	no clinical diseases	-	Not reported (but disease is known to occur)
	Suspected by reporting officer but presence not confirmed	(year)	Year of last occurrence

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	 KHV 1. Origin of the disease/pathogen: Hulu Sungai Tengah, Hulu Sungai Utara, Banjar Baru (South Kalimantan), Sukabumi, Kuningan (West Java) 2. Species affected: <i>Cyprinus carpio</i> 3. Clinical signs: wounded body, res spots on the body surface, gills damaged, most of the body surface flaking, haemorrhage. 4. Pathogen: Koi herpesvirus 5. Mortality rate: >50% (Hulu Sungai Tengah) ; 10-50% (Hulu Sungai Utara); <30% (Banjar Baru); low to high (Sukabumi and Kuningan) 6. Economic loss: - 7. Names of infected areas: Hulu Sungai Tengah, Hulu Sungai Utara, Banjar Baru (South Kalimantan), Sukabumi, Kuningan (West Java) 8. Preventive/control measures: eradication of infected fish, use of vaccine, supplementation with vitamins. 9. Laboratory confirmation: Freshwater Aquaculture Development Center Mandiangin Laboratory, Main Center Freshwater Aquaculture Development Sukabumi Laboratory. 10. Publications : not published.
2	 GIV 1. Origin of the disease/pathogen: Mariculture Development Center Lombok 2. Species affected: Epinephelus fuscoguttatus 3. Clinical signs: decreased appetite, melanosis. 4. Pathogen: Iridovirus 5. Mortality rate: 30-60% (West Lombok); 6. Economic loss: - 7. Names of infected areas: West Lombok 8. Preventive/control measures: reduce density, vaccination; 9. Laboratory confirmation: Mariculture Development Center Lombok Laboratory; 10. Publications: not published.

3	 VER Origin of the disease/pathogen: Puhawang island (Lampung Province); Species affected: Cromileptes altivelis, E. fuscoguttatus Clinical signs: fin rot; Pathogen: VNN; Mortality rate: <30% Economic loss: - Names of infected areas: Pesawaran (Lampung Province); Preventive/control measures: quarantine of sick fish, provision of good feed, vitamins and immunostimulants; Laboratory confirmation: Main Center Mariculture Development Lampung Laboratory; Publications: not published.
4	 WSD Origin of the disease/pathogen: Hanura (Lampung Province) Species affected: Litopenaeus vanname Clinical signs: - Pathogen: White spot syndrome virus, white spot baculovirus complex Mortality rate: <30%; Economic loss: - Names of infected areas: Hanura; Preventive/control measures: early harvest; Laboratory confirmation: Main Center Mariculture Development Lampung Laboratory; Publications: not published.
5	 IMN Origin of the disease/pathogen: Tanggamus (Lampung Province); Species affected: Litopenaeus vannamei Clinical signs: - Pathogen: Infectious myonecrosis virus Mortality rate: - Economic loss: - Names of infected areas: Tanggamus (Lampung Province); Preventive/control measures: early harvest; Laboratory confirmation: Main Center Mariculture Development Lampung; Publications: not published.

Country: INDONESIA

Item		Disease status ⁴	<u>a/</u>		F
DISEASES PREVALENT IN THE REGION	Month			Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	0000	0000	0000		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	+()	+()	+()	I,III	1
Non OIE-listed diseases					
8. Grouper iridoviral disease	+()	+()	+()	II	2
9. Viral encephalopathy and retinopathy	***	+()	***	III	3
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	0000	0000	0000		
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis					
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	+()	+()	+()	III	4
2. White spot disease	+()	+()	+()	III	5
3. Yellowhead disease	0000	0000	0000		
4. Infectious hypodermal and haematopoietic necrosis	+()	***	+()	III	6
5. Infectious myonecrosis	+()	***	+()	I,III	7
6.White tail disease (MrNV)	0000	0000	0000		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		
9. Monodon slow growth syndrome	0000	0000	0000		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

LISTED Finfish: Molluscs Crustace NOT LIS	ES PRESUMED EXOTIC TO THE REGION ^b BY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . s: Infection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> eans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE Channel catfish virus disease	inus.	
<u>a</u> / Please	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
+ +?	Serological evidence and/or isolation of causative agent but		
	1 1	***	No information available

Г

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

٦

Comment No.	
1	 KHV 1. Origin of the disease/pathogen: Hulu Sungai Utara, Banjar (South Kalimantan), Purwakarta, Sukabumi (West Java), Kutai Kartanegara (East Kalimantan); 2. Species affected: Cyprinus carpio, Koi 3. Clinical signs: gill rot, wound on the body surface, redness on body, haemorrahge. 4. Pathogen: Koi herpesvirus 5. Mortality rate: >70% (Hulu Sungai Utara); 6. Economic loss: Rp 5-20 million (Hulu Sungai Utara); Rp 10 million (Koi in West Java Province) 7. Names of infected areas: Hulu Sungai Utara, Banjar (South Kalimantan), Kutai Kartanegara (East Kalimantan) 8. Preventive/control measures: eradication of infected fish, use of herbal medicines, supplementation with vitamins, transfer to another pond; 9. Laboratory confirmation: Freshwater Aquaculture Development Center Mandiangin Laboratory, Main Center Freshwater Aquaculture Development Sukabumi Laboratory. 10. Publications : not published.
2	 GIV 1. Origin of the disease/pathogen: Batam, Tanjang Balai Karimun (Riau Island), South Lampung (Lampung), West Lombok (West Nusa Tenggara) 2. Species affected: Trachinotus blochii, Lates calcarifer, Oreochromis niloticus, Chromileptes altivelis 3. Clinical signs: blackened body, irregular swimming, loss of appetite, white eye ring, mass mortality. 4. Pathogen: Iridovirus; muti-infection with Vibrio sp. and Dicplectatum sp in gills. 5. Mortality rate: 1,200 pcs/month (Riau Island), >60% (West Nusa Tenggara); 6. Economic loss: Rp8 million/8000 pcs (Batam), Rp16.8 million (Tanjung Balai); 7. Names of infected areas: Batam, Tanjang Balai Karimun Berundung, South Lampung (Lampung), West Lombok (West Nusa Tenggara) 8. Preventive/control measures: Vitamins and immunostimulants; 9. Laboratory confirmation: Mariculture Development Center Lombok Laboratory; 10. Publications: not published.

3	 VER Origin of the disease/pathogen: Teluk Harun (Lampung Province); Species affected: <i>Epinephelus fuscoguttatus</i> Clinical signs: haemorrhage at the bottom of the mouth, anal fin rot, mutifocal necrosis of liver, swollen spleen and kidney, lesions and ulcers in the mouth; Pathogen: VNN; Mortality rate: 20% Economic loss: - Names of infected areas: Teluk Harun (Lampung Province); Preventive/control measures: quarantine immersion in freshwater, vitamins; Laboratory confirmation: Main Center Mariculture Development Lampung Laboratory; Publications: not published.
4	 TS 1. Origin of the disease/pathogen: Tangerang (Banten), Pinrang, Maros (South Sulawesi), South Lampung (Lampung Province); 2. Species affected: Litopenaeus vannamei 3. Clinical signs: - 4. Pathogen: TSV; 5. Mortality rate: <30% (Banten, South Sulawesi) 6. Economic loss: - 7. Names of infected areas: Tangerang (Banten), Pinrang, Maros (South Sulawesi), South Lampung (Lampung Province); 8. Preventive/control measures: Biosecurity; 9. Laboratory confirmation: Center of Fish Disease and Environment Investigation Banten Laboratory, Brackishwater Aquaculture Development Center Takalaar Laboratory, Main Center Mariculture Development Lampung Laboratory; 10. Publications: not published.
5	 WSD Origin of the disease/pathogen: Tangerang, Pandeglang (Banten), South Lampung (Lampung Province), Kota Bara (South Kalimantan), West Sumbawa (West Nusa Tenggara) Species affected: Litopenaeus vannamei, Penaeus monodon Clinical signs: soft carapace, white spot on carapace, shrimp becoming weak and swimming on the surface of water, mass mortality; Pathogen: WSSV Mortality rate: <30% (Banten), >70% (South Kalimantan), 100% (Central Java), >80% (West Nusa Tenggara); Economic loss: - Names of infected areas: Tangerang, Pandeglang (Banten), South Lampung (Lampung Province), Kota Bara (South Kalimantan), West Sumbawa (West Nusa Tenggara) Preventive/control measures: Vitamin C, immunostimulant, early harvest; Laboratory confirmation: Center of Fish Disease and Environment Investigation Banten Laboratory, Brackishwater Aquaculture Development Center Takalaar Laboratory, Main Center Mariculture Development Lampung Laboratory, Freshwater Aquaculture Development Center Mandiangin Laboratory, Main Center Brackishwater Development Jepara Laboratory, Mariculture Development Center Lombok Laboratory;

6	 IHHN Origin of the disease/pathogen: Serang (Banten); Species affected: Litopenaeus vannamei Clinical signs: white spot on tail, dwarfing Pathogen: IHHNV Mortality rate: <30% Serang (Banten and Central Java) Economic loss: - Names of infected areas: Serang (Banten), Jepara, Pati (Central Java); Preventive/control measures: biosecurity, disinfection of egg and larva; Laboratory confirmation: Center of Fish Diseases and Environment Investigation Banten Laboratory, Main Center Brackishwater Development Jepara Laboratory; Publications: not published.
7	 IMN Origin of the disease/pathogen: Tangerang (Banten), South Lampung (Lampung), Probolinggo (East Java); Species affected: Litopenaeus vannamei Clinical signs: shrimp in acute phase of IMN present focal to extensive white necrotic areas in striated (skeletal) muscles, especially in the distal abdominal segments and tail fan, becoming necrotic and reddened Pathogen: IMNV Mortality rate: <30% (Banten) Economic loss: - Names of infected areas: Berundung and Padang Cermin (Lampung Province); Preventive/control measures: early harvest, disinfection, good management; Laboratory confirmation: Center of Fish Diseases and Environment Investigation Banten Laboratory, Main Center Mariculture Development Lampung Labortory, Main Center Situbondo Laboratory; Publications: not published.

Country: IRAN

Item		Disease status ⁴	<u>1/</u>		<u></u>
DISEASES PREVALENT IN THE REGION	Month		Level of	Epidemiological comment	
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases	5	0	1		
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	-	-	-		
3. Spring viraemia of carp	-	-	-		
4. Viral haemorrhagic septicaemia	-	-	-		
5. Epizootic ulcerative syndrome	0000	0000	0000		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8. Grouper iridoviral disease	***	***	***		
9. Viral encephalopathy and retinopathy	0000	0000	0000		
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases		1			
1. Infection with Bonamia exitiosa	***	***	***		
2. Infection with Perkinsus olseni	***	***	***		
3. Infection with abalone herpes-like virus	***	***	***		
4. Infection with Xenohaliotis californiensis	***	***	***		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	***	***	***		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	-	-	+	III	1
3. Yellowhead disease	0000	0000	0000		
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000		
5. Infectious myonecrosis	***	***	***		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	***	***	***		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	***	***	***		
9. Monodon slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	***	***	***		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		
2. Infection with Batrachochytrium dendrobatidis	***	***	***		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

ctious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . fection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> : Crayfish plague (<i>Aphanomyces astaci</i>).	inus.	
: Cravfish plague (Aphanomyces astaci).		
the following symbols:		
	+()	Occurrence limited to certain zones
Disease reported or known to be present	***	No information available
Serological evidence and/or isolation of causative agent but	0000	Never reported
no clinical diseases	-	Not reported (but disease is known to occur)
Suspected by reporting officer but presence not confirmed	(vear)	Year of last occurrence
, ,	Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases	nnel catfish virus disease the following symbols: Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	 White Spot Disease Reported in 2 shrimp farms in Goatr region (Chabahar) in June 2013, origin unknown; Species affected – L. vannamei (45 d); Clinical signs – sudden decrease in feeding, swimming near the edge of pond, sudden mortality; Pathogen – White spot syndrome virus; Mortality rate – >80% (morbidity almost 100%), Economic loss – Geographic extent – Choebdeh in Khozestan Province; Control measures – both farms were disinfected with 40 ppm Calcium Chloride, all infected shrimps were eradicated; Laboratory confirmation – nested-PCR (National Shrimp Laboratory, Boushehr); Publications – None.

Country: JAPAN

Item		Disease status ^a	<u>v/</u>		<u>.</u>
DISEASES PREVALENT IN THE REGION	Month		Level of	Epidemiological comment	
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases		0			
1. Epizootic haematopoietic necrosis	0000	0000	0000	Ι	
2. Infectious haematopoietic necrosis	+	+	+	III	
3. Spring viraemia of carp	0000	0000	0000	Ι	
4. Viral haemorrhagic septicaemia	-	-	-	Ι	
5. Epizootic ulcerative syndrome	-	-	+	III	
6. Red seabream iridoviral disease	+	+	+	III	
7. Koi herpesvirus disease	+	+	+	III	
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000	Ι	
9. Viral encephalopathy and retinopathy	-	+	+	III	
10.Enteric septicaemia of catfish	-	-	-	Ι	
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000	Ι	
2. Infection with Perkinsus olseni	-	-	-	Ι	
3. Infection with abalone herpes-like virus	0000	0000	0000	Ι	
4. Infection with Xenohaliotis californiensis	+?()	+?()	-	III	1
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	-	-	-	Ι	
6. Acute viral necrosis (in scallops)	0000	0000	0000	Ι	
7. Akoya oyster disease	-	-	-	Ι	
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000	Ι	
2. White spot disease	+	+	+	III	
3. Yellowhead disease	0000	0000	0000	Ι	
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000	Ι	
5. Infectious myonecrosis	0000	0000	0000	Ι	
6. White tail disease (MrNV)	0000	0000	0000	Ι	
7. Necrotising hepatopancreatitis	0000	0000	0000	Ι	
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000	Ι	
9. Monodon slow growth syndrome	0000	0000	0000	Ι	
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000	Ι	
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	-	-	-	Ι	
2. Infection with Batrachochytrium dendrobatidis	-	-	-	Ι	
ANY OTHER DISEASES OF IMPORTANCE					

	SES PRESUMED EXOTIC TO THE REGION ^b O BY THE OIE		
	Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . s: Infection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i>	inus.	
	ceans: Crayfish plague (<i>Aphanomyces astaci</i>).		
	ISTED BY THE OIE		
nfish:	Channel catfish virus disease		
Pleas	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	***	No information available
+?	Serological evidence and/or isolation of causative agent but	0000	Never reported
	no clinical diseases	-	Not reported (but disease is known to occur)
	Suspected by reporting officer but presence not confirmed	(year)	Year of last occurrence
?		· · · ·	

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	DNA of <i>Xenohaliotis californiensis</i> was detected by PCR from <i>Haliotis diversicolor aquatilis</i> (Kanagawa Prefecture) and <i>H. diversicolor diversicolor</i> (Kagoshima Prefecture) without any clinical signs. Diagnosis was made by the National Research Institute of Aquaculture, Fishery Research Agency. Stamping out and disinfection of infected premises were implemented in response to the outbreak.
2	
3	

Country: KOREA, REPUBLIC OF Period: July - September 2013

Itam		Disease status ²	a/		L
Item DISEASES PREVALENT IN THE REGION		Month	_	Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases	<i>v</i> ary	1148400			
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	-	-	-	III	
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	-	-	-	III	
5. Epizootic ulcerative syndrome	0000	0000	0000		
6. Red seabream iridoviral disease	-	+	+	III	1
7. Koi herpesvirus disease	-	-	-	III	
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000		
9. Viral encephalopathy and retinopathy	-	-	-	III	
10.Enteric septicaemia of catfish	0000	0000	0000		1
MOLLUSC DISEASES			1		1
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	0000	0000	0000		
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	-	-	-	III	
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	+	-	-	III	2
3. Yellowhead disease	0000	0000	0000		
4. Infectious hypodermal and haematopoietic necrosis	-	-	-	III	
5. Infectious myonecrosis	0000	0000	0000		
6.White tail disease (MrNV)	0000	0000	0000		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		1
9. Monodon slow growth syndrome	0000	0000	0000		1
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES					1
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	?	?	?		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

	DBY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
lollusc	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus mar	rinus.	
	eans: Crayfish plague (<i>Aphanomyces astaci</i>).		
	STED BY THE OIE Channel catfish virus disease		
	channel cathsii virus disease		
Pleas	e use the following symbols:		
Tieus		+()	Occurrence limited to certain zones
	Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
+			
	Disease reported or known to be present	***	No information available

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	Red sea bream iridovirus (RSIV) was detected in cultured mullet (<i>Chelon haematocheilus</i>) and starry flounder (<i>Platichthys stellatus</i>) in Sachen of Gyeongsangnam-do and Gangneung Gangwon-do, respectively. No clinical signs and mortality were shown. The confirmative diagnosis was performed by National Fisheries Research and Development Institute, Aqua-life Disease Control Division. The standstill of RSIV-detected fish was declared for control.
2	White spot disease virus was detected from whiteleg shrimp (<i>Litopenaeus vanname</i>) in 2 farms in Taean-gun, Chungcheongnam-do. No clinical signs and mortality were observed. The confirmative diagnosis was performed by the National Fisheries Research and Development Institute, Aqua-life Disease Control Division. The standstill of WSDV-detected whiteleg shrimp was declared for control.
3	

Country: LAO PDR

Item		Disease status	<u>1/</u>		_
DISEASES PREVALENT IN THE REGION		Month		Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases	<i>c</i> j	8	2 · p · · · · · ·		
1. Epizootic haematopoietic necrosis	***	***	***		
2. Infectious haematopoietic necrosis	***	***	***		
3. Spring viraemia of carp	***	***	***		
4. Viral haemorrhagic septicaemia	***	***	***		
5. Epizootic ulcerative syndrome	***	***	***		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	***	***	***		
Non OIE-listed diseases					
8. Grouper iridoviral disease	***	***	***		
9. Viral encephalopathy and retinopathy	***	***	***		
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with <i>Bonamia exitiosa</i>	***	***	***		
2. Infection with <i>Perkinsus olseni</i>	***	***	***		
3. Infection with abalone herpes-like virus	***	***	***		
Non OIE-listed diseases					
4. Infection with <i>Marteilioides chungmuensis</i>	***	***	***		
5. Acute viral necrosis (in scallops)	***	***	***		
6. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	***	***	***		
2. White spot disease	***	***	***		
3. Yellowhead disease	***	***	***		
4. Infectious hypodermal and haematopoietic necrosis	***	***	***		
5. Infectious myonecrosis	***	***	***		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	***	***	***		
Non OIE-listed diseases					
8. <i>Monodon</i> slow growth syndrome	***	***	***		
9. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	***	***	***		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		
2. Infection with <i>Batrachochytrium dendrobatidis</i>	***	***	***		
ANY OTHER DISEASES OF IMPORTANCE					
1.					
2.					

nfish: ollusce ustace	BY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . E: Infection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> cans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE	rinus; Xenohalio	tis californiensis.
fich	Channel catfish virus disease		
	e use the following symbols:		
Please		+() ***	Occurrence limited to certain zones
Please +	use the following symbols: Disease reported or known to be present Serological evidence and/or isolation of causative agent but	***	No information available
Please	Disease reported or known to be present		

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	
2	
3	

Country: MALAYSIA

Item		Disease status ⁴	<u>ı/</u>		Epidemiological
DISEASES PREVALENT IN THE REGION		Month		Level of diagnosis	comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000	I,II,III	
3. Spring viraemia of carp	0000	0000	0000	I,II,III	1
4. Viral haemorrhagic septicaemia	0000	0000	0000	I,II,III	
5. Epizootic ulcerative syndrome	(1986)	(1986)	(1986)	I.II	
6. Red seabream iridoviral disease	-	-	-	I,II,III	
7. Koi herpesvirus disease	-	-	-	I,II,III	2
Non OIE-listed diseases					
8. Grouper iridoviral disease	-	-	-	III	3
9. Viral encephalopathy and retinopathy	-	-	-	III	4
10.Enteric septicaemia of catfish	0000	0000	0000		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with <i>Perkinsus olseni</i>	0000	0000	0000	III	
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis					
Non OIE-listed diseases					
5. Infection with <i>Marteilioides chungmuensis</i>	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	-	-	-	I,III	5
2. White spot disease	-	+	+	I,III	6
3. Yellowhead disease	-	-	-	I,III	7
4. Infectious hypodermal and haematopoietic necrosis	+	+	-	I,III	8
5. Infectious myonecrosis	-	-	-	III	9
6.White tail disease (MrNV)	-	-	-	III	10
7. Necrotising hepatopancreatitis	-	-	-	III	11
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	0000	0000	0000		
9. Monodon slow growth syndrome	-	-	-		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	+	-	-	I,II,III	13
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	-	-	-		
2. Infection with <i>Batrachochytrium dendrobatidis</i>	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					
1. Hepatopancreatic parvo virus disease	-	+	+	III	12

infish: lollusc:	 DY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i>. s: Infection with <i>Bonamia ostreae</i>; <i>Marteilia refringens</i>; <i>Perkinsus marteilia</i> 	inus.	
OT LI	eans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE Channel catfish virus disease		
	e use the following symbols:		
1 ICasy	e use the following symbols.	+()	Occurrence limited to certain zones
+	Disease reported or known to be present	***	No information available
+ +?	Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases		No information available Never reported Not reported (but disease is known to occur)

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	Spring viraemia of carp 1. No positive cases detected (PCR) during DoF active surveillance programme
2	Koi herpesvirus disease 1. One positive case was detected (PCR) during DoF active surveillance programme
3	 Grouper Iridoviral disease (GIV) 1. Imported grouper fish fry (sample from Selangor) and barramundi fish fry were positive for GIV tested in Fisheries Biosecurity Penang for disease surveillance (herewith referred to Megalocytivirus).
4	 Viral encephalopathy and retinopathy 1. Imported grouper fish fry (sample from Selangor) were positive for VNN tested in Fisheries Biosecurity KLIA Selangor for disease surveillance.
5	 Taura syndrome virus (TSV) (<i>Penaeus monodon, Litopenaeus vannamei</i>) 1. TSV was not detected in all the 26 samples of postlarvae sent to Private laboratory for routine and monitoring purposes. 2. No positive on reported cases detected by PCR although active surveillance was conducted by DoF in West and East Malaysia.

 White Spot Syndrome Virus (WSSV) Three cases of WSSV in <i>P. vannamei</i> were detected out of 147 samples sent to Private Laboratory for routine and monitoring purposes. It involved juvenile and postlarvae of <i>P. monodon</i> and <i>P. vannamei</i>. No positive cases detected by PCR although active surveillance was conducted by DOF in West and East Malaysia.
 Yellow head disease (YHV) (P. monodon, Litopenaeus vannamei) 1. YHV was not detected in all the 18 samples sent to private laboratory for routine and monitoring purposes. 2. No positive cases detected (PCR) although active surveillance was conducted by DoF in East Malaysia
 Infectious hypodermal and haematopoietic necrosis virus (IHHNV) (Macrobrachium rosenbergii, P. monodon, Litopenaeus vannamei) 1. No IHHNV was detected 25 L. vannamei postlarval samples sent to private laboratory for routine and monitoring purposes. 2. One (1) positive case detected by PCR through active surveillance conducted by DoF in West and East Malaysia.
 Infectious Myonecrosis (IMNV) IMNV was not detected in all 41 samples of <i>Penaeus monodon</i> and <i>Litopenaeus vannamei</i> postlarvae and juveniles sent to private laboratory for routine and monitoring purposes. No positive on reported cases detected by PCR, although active surveillance was conducted by DoF in West and East Malaysia.
Macrobrachium rosenbergii Nodavirus (MrNV) 1. No samples were tested for MrNV.
Necrotising hepatopancreatitis (NHPB) 1. No samples were tested for NHPB.
 Hepatopancreatic parvo virus disease (HPV) (P. monodon, Litopenaeus vannamei) 1. 13 out of 46 postlarval samples (P. monodon) were tested positive for HPV by private laboratory for routine and monitoring purposes.

	Acute hepatopancreatic necrosis syndrome (AHPNS)
13	 Early AHPNS on 16 samples from outbreak cases in 6 ponds in Sarawak; tested positive by National Fish Disease Research Institute (NaFisH), Penang in July. No positive cases in 11 samples from 2 pond outbreak cases in Setiu, Terengganu in July. No positive cases in 20 samples from 4 pond outbreak cases 11 samples from 2 pond outbreak cases in Setiu, Terengganu.

Country: MYANMAR

14.		D: a/			
Item DISEASES PREVALENT IN THE REGION		Disease status ^{a/} Month	Level of	Epidemiological	
FINFISH DISEASES	July	August	September	diagnosis	comment numbers
OIE-listed diseases	<i>tury</i>	1100000	September		
1. Epizootic haematopoietic necrosis	***	***	***		
2. Infectious haematopoietic necrosis	***	***	***		
3. Spring viraemia of carp	***	***	***		
4. Viral haemorrhagic septicaemia	***	***	***		
5. Epizootic ulcerative syndrome	***	***	***		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease					
Non OIE-listed diseases					
8. Grouper iridoviral disease	***	***	***		
9. Viral encephalopathy and retinopathy	***	***	***		
10.Enteric septicaemia of catfish	***	***	***		1
MOLLUSC DISEASES					1
OIE-listed diseases	/	/	/		1
1. Infection with Bonamia exitiosa	/	/	/		1
2. Infection with <i>Perkinsus olseni</i>	/	/	/		1
3. Infection with abalone herpes-like virus	/	/	/		
4. Infection with Xenohaliotis californiensis	/				
Non OIE-listed diseases	/	/			
5. Infection with Marteilioides chungmuensis	/	/	/		
6. Acute viral necrosis (in scallops)	/	/	/		
7. Akoya oyster disease					
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	-	-	-	III	1
2. White spot disease	-	-	-	III	
3. Yellowhead disease	-	-	-	III	
4. Infectious hypodermal and haematopoietic necrosis	-	-	-	III	
5. Infectious myonecrosis	***	***	***		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	***	***	***		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (Panulirus spp.)	***	***	***		
9. Monodon slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)					1
AMPHIBIAN DISEASES					1
OIE-listed diseases					
1. Infection with Ranavirus					
2. Infection with Batrachochytrium dendrobatidis					
ANY OTHER DISEASES OF IMPORTANCE					1
1.					1

	DATION OF A STATE OF 		
	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus mar	rinus.	
	eans: Crayfish plague (<i>Aphanomyces astaci</i>).		
	STED BY THE OIE Channel catfish virus disease		
iiii3ii. V			
Please	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	***	No information available
+?	Serological evidence and/or isolation of causative agent but	0000	Never reported
	no clinical diseases	-	Not reported (but disease is known to occur)
	Suspected by reporting officer but presence not confirmed	(year)	Year of last occurrence

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	During this period, we have received 7 samples of shrimps (2 frozen and 5 live shrimps for export) for testing for TSV, WSSV and IHHNV. All samples were found negative for the viruses.
2	
3	

Country: NEPAL

L. L.	Item Disease status $\frac{a'}{a}$				
Item DISEASES PREVALENT IN THE REGION		Disease status Month	Level of	Epidemiological	
FINFISH DISEASES	July	August	September	diagnosis	comment numbers
OIE-listed diseases	varj	Tagast	September		
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	-	-	-		1
6. Red seabream iridoviral disease	0000	0000	0000		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8.Grouper iridoviral disease					
9.Viral encephalopathy and retinopathy					
10.Enteric septicaemia of catfish					
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	***	***	***		
2. Infection with <i>Perkinsus olseni</i>	***	***	***		
3. Infection with abalone herpes-like virus	***	***	***		
4. Infection with <i>Xenohaliotis californiensis</i>					
Non OIE-listed diseases					
5. Infection with <i>Marteilioides chungmuensis</i>	***	***	***		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	***	***	***		
2. White spot disease	***	***	***		
3. Yellowhead disease	***	***	***		
4. Infectious hypodermal and haematopoietic necrosis	***	***	***		
5. Infectious myonecrosis	***	***	***		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	***	***	***		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	***	***	***		
9. Monodon slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)					
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		
2. Infection with Batrachochytrium dendrobatidis	***	***	***		
ANY OTHER DISEASES OF IMPORTANCE					
1.					

ctious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . fection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> : Crayfish plague (<i>Aphanomyces astaci</i>).	inus.	
: Cravfish plague (Aphanomyces astaci).		
the following symbols:		
	+()	Occurrence limited to certain zones
Disease reported or known to be present	***	No information available
Serological evidence and/or isolation of causative agent but	0000	Never reported
no clinical diseases	-	Not reported (but disease is known to occur)
Suspected by reporting officer but presence not confirmed	(vear)	Year of last occurrence
, ,	Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases	nnel catfish virus disease the following symbols: Disease reported or known to be present Serological evidence and/or isolation of causative agent but no clinical diseases

1. Epidemiological comments:

(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (unusual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc). and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	 Epizootic ulcerative syndrome Seen in Chitawan and Rupandehi districts in September; Species affected – mostly Chinese carps (200-1000 g); Clinical signs – scale removal, haemorrhage on finbase and operculum edge; Pathogen –; Mortality rate –, Economic loss – 1 ton in Chitawan and 1.5 tons in Rupandehi Geographic extent – 200-700 g size of Indian carps in 25 ha. Ponds; 75-200 g catfishes in 100 small kitchen ponds; Control measures – Potassium permanganate @ 500kg/ha for 7-0 days, and 500 kg/ha lime; Laboratory confirmation –; Publications – None.
2	
3	

2. New aquatic animal health regulations introduced within past six months (with effective date):

Country: <u>PHILIPPINES</u>

Period: July - September 2013

Item		Disease status a	<u>/</u>		
DISEASES PREVALENT IN THE REGION		Month		Level of	Epidemiological comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases		<u> </u>			
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	- (2002)	- (2002)	- (2002)	Ι	1
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	0000	0000	0000	III	
Non OIE-listed diseases					
8. Grouper iridoviral disease	- (2008)	- (2008)	- (2008)	III	2
9. Viral encephalopathy and retinopathy	+	+	+	III	3
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	0000	0000	0000		
3. Infection with abalone herpes-like virus	***	***	***		
4. Infection with Xenohaliotis californiensis	***	***	***		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000	III	4
2. White spot disease	+	+	+	III	5
3. Yellowhead disease	- (1999)	- (1999)	- (1999)	III	6
4. Infectious hypodermal and haematopoietic necrosis	+	+	+	III	7
5. Infectious myonecrosis	0000	0000	0000	III	8
6.White tail disease (MrNV)	0000	0000	0000	III	9
7. Necrotising hepatopancreatitis	0000	0000	0000	III	10
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (Panulirus spp.)	***	***	***		
9. Monodon slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		
2. Infection with Batrachochytrium dendrobatidis	***	***	***		
ANY OTHER DISEASES OF IMPORTANCE					
1. Monodon baculovirus (MBV)	-	-	-	III	

	DBY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
lollusc	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus mar	rinus.	
	eans: Crayfish plague (<i>Aphanomyces astaci</i>).		
	STED BY THE OIE Channel catfish virus disease		
	chamer catrisii virus disease		
Pleas	e use the following symbols:		
Tieus		+()	Occurrence limited to certain zones
	Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
+			
	Disease reported or known to be present	***	No information available

1. Epidemiological comments:

Comment No.	
1	Sixty five (65) pieces of eels (<i>Anguilla spp.</i>) were negative for Epizootic Ulcerative Syndrome by gross morphological examination. Samples were from Laguna and Cotabato City. Examination was conducted by the Bureau of Fisheries and Aquatic Resources (BFAR) Central Office Fish Health Laboratory.
2	Four (4) samples - (3 grouper, 1 snapper) were analyzed using Polymerase Chain Reaction (PCR) test. All samples showed negative results for Iridoviral Disease. The samples were collected from Sarangani Province, Sorsogon and Nueva Ecija. Examination was conducted by BFAR Central Office Fish Health Laboratory and Southeast Asian Fisheries and Development Center/ Aquaculture Department (SEAFDEC/AQD) Laboratory.
3	Twenty (20) samples – (6 grouper, 6 pompano, 5 snapper, 2 siganid and 1 seabass) were analyzed using PCR test. Twelve (12) samples- (6 Grouper, 4 Pompano and 2 Saganid) showed positive result for Viral Encephalopathy and Retinopathy. The positive samples were collected from Sorsogon, Guimaras and Iloilo. Examination was conducted by the BFAR Central Office and SEAFDEC/AQD Laboratories.
4	One hundred sixty five (165) samples - (137 <i>Penaeus vannamei</i> ,27 <i>Penaeus monodon</i> and 1 <i>M.rosenbergii</i>) of different stages (brood stock, adult, fry, juvenile) were analyzed using PCR test. All samples showed negative results for Taura Syndrome. The samples were collected from Bohol, Sorsogon, Zamboanga City, Sarangani Province, Surigao del Sur, Cotabato City, General Santos City, Quezon, Batangas, Samar,Ormoc City, Tacloban, Iloilo, Rizal, Cebu, Misamis Occidental, Lanao del Norte, Davao Oriental, Aurora, Bataan and Dapitan. Other samples were imported from Hawaii and Florida, USA. Examinations were conducted by BFAR Central Office, BFAR Region III, BFRA Region VI, BFAR Region VII and BFAR Region VIII and SEAFDEC/AQD Laboratories.

5	Four hundred ninety-five (495) samples of <i>P.vannamei, P.monodon, S.serrata,</i> artemia and crabs of different stages (fry, juvenile, adult and brood stock) were tested using PCR. Twenty-one - (10 <i>P.vannamei,8 P.monodon</i> , 3 crabs) were positive for White Spot Syndrome Virus. The positive samples were from Bohol, Cotabato City, Sorsogon, Samar, Sarangani, Camarines Sur, General Santos and Cebu. Examinations were conducted by BFAR Central Office, BFAR Region III, BFAR Region VI, BFAR Region VII, BFAR Region VII, Negros Prawn Producers Cooperative and SEAFDEC/AQD Laboratories.
6	One hundred forty (140) samples - (112 <i>P.vannamei</i> , 27 <i>P.monodon</i> and 1 <i>M.rosenbergii</i>) of different stages (fry, juvenile, grow out, adult and brood stock) were tested using PCR. All samples showed negative results for Yellowhead Disease. The samples were collected from Sorsogon, Zamboanga City, Sarangani, Surigao del Sur, Cotabato City, General Santos City, Quezon Province, Zambales, Batangas, Samar, Ormoc City, Tacloban City, Iloilo, Cebu, Rizal, Bohol,Misamis Occidental, Lanao del Norte, Davao Oriental, Dapitan City and Aurora. Other samples were imported from Hawaii and Florida, USA. Examinations were conducted by BFAR Central Office, BFAR Region III, BFAR Region VI, BFAR Region VII, BFAR Region VIII and SEAFDEC/AQD Laboratories.
7	One hundred ninety-seven (197) samples - (52 <i>P.monodon</i> , 143 <i>P.vannamei</i> , 1 <i>M.rosenbergii</i> and 1 artemia) of different stages (fry, juvenile, adult, brood stock) were analyzed using the PCR Test. Eleven (7 <i>P.vannamei</i> , 4 <i>P.monodon</i> ,) showed positive result for Infectious Hypodermal and Haematopoietic Necrosis Virus. The positive samples were collected from Samar, Iloilo and Batangas. Examinations were conducted by BFAR Central Office, BFAR Region III, BFAR Region VI, BFAR Region VII, BFAR Region VIII and SEAFDEC/AQD Laboratories.
8	One hundred sixty-five (165) samples - (137 <i>P.vannamei</i> , 27 <i>P.monodon</i> , and 1 <i>M.rosenbergii</i>) of different stages (fry, juvenile, adult and bloodstock) were tested using PCR. All samples showed negative results for Infectious Myonecrosis. The samples were collected from Sorsogon, Zamboanga City, Sarangani Province, Surigao del Sur, Cotabato City, General Santos City, Quezon Province, Zambales, Batangas, Samar, Ormoc City, Tacloban City, Iloilo, Cebu, Rizal, Bohol and Bataan. Other samples were imported from Hawaii, USA. Examinations were conducted by BFAR Central Office, BFAR Region III, BFAR Region VI, BFAR Region VII, BFAR Region VIII and SEAFDEC/AQD Laboratories.
9	One (1) sample of <i>M.rosenbergii</i> was tested using PCR. The sample showed negative results for White Tail Disease. The sample was collected from Rizal. Examination was conducted by BFAR Central Office Fish Health Laboratory.
10	One hundred fourteen (114) samples - (86 <i>P.vannamei</i> ,27 <i>P.monodon</i> and 1 <i>M.rosenbergii</i>) of various stages (fry, juvenile, adult and brood stock) were tested using PCR. All samples showed negative results for Necrotising Hepatopancreatitis. The samples were collected from Sorsogon, Zamboanga City, Sarangani Province, Surigao del Sur, Cotabato City, General Santos City, Quezon Province, Zambales, Batangas, Samar, Ormoc City, Iloilo, Rizal, Cebu, Bohol, Davao del Sur and Dapitan City. Other samples were imported from Hawaii and Florida, USA. Examination was conducted by BFAR Central Office, BFAR Region III, BFAR Region VI, BFAR Region VII and BFAR Region VIII Laboratories.

2. New aquatic animal health regulations introduced within past six months (with effective date):

Country: SINGAPORE

Period: July - September 2013

Item		Disease status	<u>a/</u>		Epidemiological
DISEASES PREVALENT IN THE REGION		Month		Level of diagnosis	comment
FINFISH DISEASES	July	August	September	ulagilosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	0000	0000	0000		
6. Red seabream iridoviral disease	(2013)	(2013)	(2013)		
7. Koi herpesvirus disease	(2012)	(2012)	(2012)	III	1
Non OIE-listed diseases					
8. Grouper iridoviral disease	(2013)	+	(2013)	III	2
9. Viral encephalopathy and retinopathy	+	+	+	III	3
10.Enteric septicaemia of catfish	0000	0000	0000		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	***	***	***		
2. Infection with Perkinsus olseni	***	***	***		
3. Infection with abalone herpes-like virus	***	***	***		
4. Infection with <i>Xenohaliotis californiensis</i>	***	***	***		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	***	***	***		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	(2013)	(2013)	(2013)	III	4
3. Yellowhead disease	0000	0000	0000		
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000		
5. Infectious myonecrosis	0000	0000	0000		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	***	***	***		
9. <i>Monodon</i> slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	0000	0000	0000		
AMPHIBIAN DISEASES	0000				1
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		1
2. Infection with <i>Batrachochytrium dendrobatidis</i>	+?	+?	+?	III	5

AN	Y OTHER DISEASES OF IMPORTANCE					
1.	Infectious spleen and kidney necrosis virus (ISKNV) (marine and ornamental fish)	+	+	+	III	6
2.	Aeromonas salmonicida (in goldfish)	0000	0000	0000	III	7

	DATINE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
Iollusc	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus man	inus.	
	eans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE		
	Channel catfish virus disease		
/ Pleas	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
+	Disease reported or known to be present	***	No information available

<u>b</u>/ If there is suspicion or confirmation of any of these diseases, they must be reported immediately, because the region is considered free of these diseases

1. Epidemiological comments:

Comment No.	
1	Koi herpesvirus (KHV) was not detected in one batche of koi submitted on 26 th September with slight reddening of the body and lethargy by real-time PCR. Anti-KHV antibodies were not detected in serum of these koi by serology. Monogenean parasites were detected in on skin and gills and virus isolation for SCVC is in progress. 31 batches of imported and locally farmed ornamental koi from surveillance programs and voluntary submission tested negative by real-time PCR.
2	Singapore grouper iridovirus (SGIV) was detected by PCR in 1 out of 2 batches of diseased imported pearl grouper (<i>E. lanceolatus</i> x <i>E. fuscoguttatus</i>) fingerlings in August. Infection with VNNV was confirmed by histopathological findings and qPCR in these fish (see comment 3). SGIV was not detected in 1 batch of local pearl grouper.
3	Viral nervous necrosis virus (VNNV) was detected in a batch of locally bred pearl grouper in July, which were also infected with ISKNV (see comment 6). In August, VNNV was detected in 5 batches of local and imported pearl grouper from land-based hatcheries and floating net cage farms, and one batch of hybrid pompano from a land-based fish hatchery. VNNV was also detected in 1 out of 2 batches of yellowtail scad submitted from a private aquaria in September. All detections of VNNV were confirmed by histopathological examination and qPCR.

4	White spot syndrome virus (WSSV) was not detected in .24 batches of ornamental crustaceans submitted for targeted surveillance programs, 1 voluntary submission by an exporter, and in120 <i>Litopenaeus vannamei</i> submitted from a local broodstock farm this quarter.
5	112 frog swab samples were submitted from June to September for validation of the <i>Batrachochytrium dendrobatidis</i> (Bd) real-time PCR protocol. Reactors and negative samples will be submitted to a diagnostic laboratory for confirmation. Surveillance in imported ornamental frogs, and imported and locally farmed frogs will start at the end of November.
6	Infectious spleen and kidney necrosis virus (ISKNV) was detected in a batch of diseased pearl grouper in July which were also infected with VNNV (comment 3), and 1 batch of diseased gourami in August. <i>Edwardsiella</i> sp. and <i>Plesiomonas shigelloides</i> was isolated from the gourami samples. One batch of imported pomapano tested positive for ISKNV. All detections of ISKNV were confirmed by histopathologial examination and PCR.
5	<i>Aeromonas salmonicida</i> was not detected this quarter in all 15 batches of goldfish submitted under a targeted surveillance program for goldfish exported to Australia.

2. New aquatic animal health regulations introduced within past six months (with effective date):

Country: SRI LANKA

Period: July - September 2013

Item		Disease status	<u>a/</u>		Epidemiological
DISEASES PREVALENT IN THE REGION		Month		Level of	comment
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000	III	1
2. Infectious haematopoietic necrosis	***	***	***		
3. Spring viraemia of carp	0000	0000	0000	III	2
4. Viral haemorrhagic septicaemia	0000	0000	0000	III	3
5. Epizootic ulcerative syndrome	***	***	***		
6. Red seabream iridoviral disease	***	***	***		
7. Koi herpesvirus disease	0000	0000	0000	III	4
Non OIE-listed diseases					
8.Grouper iridoviral disease	***	***	***		
9. Viral encephalopathy and retinopathy	***	***	***		
10.Enteric septicaemia of catfish	***	***	***		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	***	***	***		
2. Infection with Perkinsus olseni	***	***	***		
3. Infection with abalone herpes-like virus	***	***	***		
4. Infection with Xenohaliotis californiensis					
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	***	***	***		
6. Acute viral necrosis (in scallops)	***	***	***		
7. Akoya oyster disease	***	***	***		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	***	***	***		
2. White spot disease	+()	+()	+()	III	5
3. Yellowhead disease	+()	?()	?()	III	6
4. Infectious hypodermal and haematopoietic necrosis	?()	?()	?()	III	7
5. Infectious myonecrosis	***	***	***		
6.White tail disease (MrNV)	***	***	***		
7. Necrotising hepatopancreatitis	***	***	***		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	***	***	***		
9. Monodon slow growth syndrome	***	***	***		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	***	***	***		
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	***	***	***		
2. Infection with Batrachochytrium dendrobatidis	***	***	***		

ANY OTHER DISEASES OF IMPORTANCE					
1. Laem Singh virus (LSV)	?()	?()	?()	III	8
2. Monodon Baculovirus (MBV)	?()	?()	+()	III	9

	SES PRESUMED EXOTIC TO THE REGION ^b D BY THE OIE		
	Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> .		
	s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus ma	rinus.	
	ceans: Crayfish plague (Aphanomyces astaci).		
OT L	ISTED BY THE OIE		
Finfish :	Channel catfish virus disease		
Finfish	Channel catfish virus disease		
	Channel catfish virus disease se use the following symbols:		
		+()	Occurrence limited to certain zones
	be use the following symbols: Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
n∕ Pleas	se use the following symbols:		
<u>a</u> / Pleas +	be use the following symbols: Disease reported or known to be present	***	No information available

 \underline{b} / If there is suspicion or confirmation of any of these diseases, they must be reported immediately, because the region is considered free of these diseases

1. Epidemiological comments:

Comment No.	
1	At the Central Veterinary Investigation Centre (CVIC), PCR has been developed for EHN. Samples were not tested for this reporting period
2	68 samples (22 carps, 8 koi and 38 guppies) were tested by PCR for SVC at CVIC. All the samples gave negative result. The samples were collected from aquarium of western and north western provinces, and these were from export samples.
3	PCR has been developed for VHS at CVIC, and no samples were tested during this reporting period.
4	A total of 23 carp samples were tested for koi herpesvirus at the Centre for Aquatic Disease Diagnosis and Research (CADDAR). These samples were received from western province and other samples were from farmers at north central province, and all the samples were negative.
5	865 samples of <i>P. monodon</i> were tested by PCR for WSSV, and 148 samples were found positive. The testing has been carried out in the Laboratories of National Aquatic Research Agency (NARA) and National Aquatic Development Authority (NAQDA).
6	10 samples of <i>P. monodon</i> were tested by PCR for YHV in the laboratory of NARA during this reporting period. One sample gave positive result.

7	13 samples of <i>P. monodon</i> were tested by PCR for IHHNV during this reporting period, and no sample was found positive. Test was carried out in the laboratory of NARA.
8	11 samples of <i>P. monodon</i> were tested by PCR for LSV during this reporting period. All samples gave negative result. Test was carried out in the laboratory of NARA.
9	10 of 65 samples of <i>P. monodon</i> gave positive result for MBV in the month of September. PCR method was used for testing at the laboratories of NARA and NAQDA.

2. New aquatic animal health regulations introduced within past six months (with effective date):

Country: VIETNAM

Period: <u>April - June 2013</u>

Item		Disease status ^{a/}			Enidamialagiaal
DISEASES PREVALENT IN THE REGION	Month			Level of	Epidemiological comment
FINFISH DISEASES	April	May	June	diagnosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	-	-	-		
6. Red seabream iridoviral disease	0000	0000	0000		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000		
9. Viral encephalopathy and retinopathy	0000	0000	0000		
10.Enteric septicaemia of catfish	-	-	-		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	+()	-	-	I,III	1
3. Infection with abalone herpes-like virus	0000	0000	0000		
4. Infection with Xenohaliotis californiensis	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	+	+	+	I,II,III	2
3. Yellowhead disease	-	-	-		
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000		
5. Infectious myonecrosis	0000	0000	0000		
6.White tail disease (MrNV)	-	-	-		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (<i>Panulirus</i> spp.)	-	-	-		
9. Monodon slow growth syndrome	-	-	-		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	+	+	+	I,III	3
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					
		1			

LISTED Finfish: Mollusc Crustac NOT LI	 SES PRESUMED EXOTIC TO THE REGION^b D BY THE OIE Infectious salmon anaemia; Infection with Gyrodactylus salaris. s: Infection with Bonamia ostreae; Marteilia refringens; Perkinsus mareans: Crayfish plague (Aphanomyces astaci). (STED BY THE OIE Channel catfish virus disease 	inus.	
<u>a</u> / Please	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present		
$^{+}_{+2}$	Disease reported or known to be present	***	No information available
+ +?	Serological evidence and/or isolation of causative agent but		No information available Never reported
	1 1	***	No information available

1. Epidemiological comments:

Comment No.	
1	Infection with <i>Perkinsus sp.</i> One disease outbreak in Meretrix lyrata was reported in Nghi Thiet commune, Nghi Loc district in Nghe An Province in April. The affected area was 2.5 ha. Control measure applied was disinfetion of affected area.
2	 White Spot Disease (WSD) Pathogen: White spot syndrome virus (WSSV) Species affected: Penaeus monodon and Litopenaeus vannamei (15-50 DOC) Name of affected area: reported in 20 provinces including Thai Binh, Thanh Hoa, Nghe An, Ha Tinh, Quang Tri, Quang Binh, Quang Nam, Quang Ngai, TT Huc, Binh Dinh, Phu Yen, Ho Chi Minh City, Long An, Tien Gang, Kien Giang, Ben Tre, Soc Trang, Tra Vinh, Bac Lieu and Ca Mau. Mortality rate: medium to high, 100% in some cases within 10 d. Clinical signs: lethargic or moribund shrimps accumulated at pond surface and edges, slow to erratic swimming behavior, overall body color often reddish, minute to large (0.5-2.0 mm diameter) white inclusions embedded in the cuticle, especially in the removed carapace held to light after scraping off attached tissue (not always seen) Control measures: early harvest, strict isolation of infected ponds with movement and transport controls, disinfection of infected ponds using Calcium hypochlorite (chlorine).

	Acute hepatopancreatic necrosis syndrome (AHHPNS)
	Pathogen: Vibrio parahaemolyticus with Phage A3
	Name of affected area: The disease is still affecting 12 provinces including Nam Dinh (North), Nghe An, Quang Binh, Quang Tri, Phy Yen, Ninh Thuan (Central Coast), and Ba Ria-Vung Tau, Ho Chi Minh City, Ben Tre, Tra Vinh, Bac Lieu and Ca Mau (Mekong River Delta).
5	Mortality rate: Mortality recorded at 10-45 days post stocking in both <i>P. monodon</i> and <i>L. vannamei</i> in intensive and semi-intensive farming systems was as high as 95%.
	Clinical signs: lethargy, soft and darkened shells, and mottling of the carapace; pathology appears to be limited to the hepatopancreas.
	Control measures: strict isolation of infected ponds; movement/transportation controls; use of calcium hypochlorite (Chlorine) to disinfect ponds.

2. New aquatic animal health regulations introduced within past six months (with effective date): None

Country: VIETNAM

Period: July - September 2013

Item		Disease status ²		Epidemiological comment	
DISEASES PREVALENT IN THE REGION	Month				Level of diagnosis
FINFISH DISEASES	July	August	September	diagnosis	numbers
OIE-listed diseases					
1. Epizootic haematopoietic necrosis	0000	0000	0000		
2. Infectious haematopoietic necrosis	0000	0000	0000		
3. Spring viraemia of carp	0000	0000	0000		
4. Viral haemorrhagic septicaemia	0000	0000	0000		
5. Epizootic ulcerative syndrome	-	-	-		
6. Red seabream iridoviral disease	0000	0000	0000		
7. Koi herpesvirus disease	0000	0000	0000		
Non OIE-listed diseases					
8. Grouper iridoviral disease	0000	0000	0000		
9. Viral encephalopathy and retinopathy	0000	0000	0000		
10.Enteric septicaemia of catfish	-	-	-		
MOLLUSC DISEASES					
OIE-listed diseases					
1. Infection with Bonamia exitiosa	0000	0000	0000		
2. Infection with Perkinsus olseni	0000	0000	0000		
3. Infection with abalone herpes-like virus	-	-	-		
4. Infection with Xenohaliotis californiensis	0000	0000	0000		
Non OIE-listed diseases					
5. Infection with Marteilioides chungmuensis	0000	0000	0000		
6. Acute viral necrosis (in scallops)	0000	0000	0000		
7. Akoya oyster disease	0000	0000	0000		
CRUSTACEAN DISEASES					
OIE-listed diseases					
1. Taura syndrome	0000	0000	0000		
2. White spot disease	+	+	+	I,II,III	1
3. Yellowhead disease	-	-	-		
4. Infectious hypodermal and haematopoietic necrosis	0000	0000	0000		
5. Infectious myonecrosis	0000	0000	0000		
6.White tail disease (MrNV)	-	-	-		
7. Necrotising hepatopancreatitis	0000	0000	0000		
Non OIE-listed diseases					
8. Milky haemolymph disease of spiny lobster (Panulirus spp.)	-	-	-		
9. <i>Monodon</i> slow growth syndrome	-	-	-		
10. Acute hepatopancreatic necrosis syndrome (AHPNS)	+	+	+	I,II	2
AMPHIBIAN DISEASES					
OIE-listed diseases					
1. Infection with Ranavirus	0000	0000	0000		
2. Infection with Batrachochytrium dendrobatidis	0000	0000	0000		
ANY OTHER DISEASES OF IMPORTANCE					1
			1		

LISTED Finfish: Molluscs Crustace NOT LIS	ES PRESUMED EXOTIC TO THE REGION ^b BY THE OIE Infectious salmon anaemia; Infection with <i>Gyrodactylus salaris</i> . s: Infection with <i>Bonamia ostreae</i> ; <i>Marteilia refringens</i> ; <i>Perkinsus mar</i> eans: Crayfish plague (<i>Aphanomyces astaci</i>). STED BY THE OIE Channel catfish virus disease	inus.	
<u>a</u> / Please	e use the following symbols:		
		+()	Occurrence limited to certain zones
+	Disease reported or known to be present	+() ***	Occurrence limited to certain zones No information available
+ +?	Serological evidence and/or isolation of causative agent but		
	1 1	***	No information available

1. Epidemiological comments:

Comment No.	
1	 White Spot Disease (WSD) Pathogen: White spot syndrome virus (WSSV) Species affected: Penaeus monodon and Litopenaeus vannamei (15-40 DOC) Name of affected area: reported in 14 provinces including Nam Dinh, Nghe An, Ha Tinh, Quang Tri, Quang Ngai, Ninh Thuan, Ho Chi Minh City, Tien Giang, Ben Tre, Tra Vinh, Kien Gang, Soc Trang, Bac Lieu and Ca Mau. Mortality rate: medium to high, 100% in some cases within 10 days. Clinical signs: lethargic or moribund shrimps accumulated at pond surface and edges, slow to erratic swimming behavior, overall body color often reddish, minute to large (0.5-2.0 mm diameter) white inclusions embedded in the cuticle, especially in the removed carapace held to light after scraping off attached tissue (not always seen) Control measures: early harvest, strict isolation of infected ponds with movement and transport controls, disinfection of infected ponds using Calcium hypochlorite (chlorine).

	Acute hepatopancreatic necrosis syndrome (AHHPNS)
	Pathogen: Vibrio parahaemolyticus with Phage A3
	Name of affected area: The disease is still affecting 11 provinces (around 1,700 ha) including Quan Ninh (North), Nghe An, Ha Tinh, Phu Yen, Ninh Thuan (Central Coast), and Ho Chi Minh City, Ben Tre, Tien Gang, Tra Vinh, Bac Lieu and Ca Mau (South; Mekong Delta).
2	Mortality rate: Mortality recorded at 10-45 days post stocking in both <i>P. monodon</i> and L. vannamei in intensive and semi-intensive farming systems was as high as 95%.
	Clinical signs: lethargy, soft and darkened shells, and mottling of the carapace; pathology appears to be limited to the hepatopancreas.
	Control measures: strict isolation of infected ponds; movement/transportation controls; use of calcium hypochlorite (Chlorine) to disinfect ponds.

2. New aquatic animal health regulations introduced within past six months (with effective date): None

List of Diseases in the Asia-Pacific Quarterly Aquatic Animal Disease Report (Beginning 2013)

1. DISEASES PREVALENT IN THE REGION		
1.1 FINFISH DISEASES		
OIE-listed diseases	Non OIE-listed diseases	
1. Epizootic haematopoietic necrosis	1.Grouper iridoviral disease	
2. Infectious haematopoietic necrosis	2. Viral encephalopathy and retinopathy	
3. Spring viraemia of carp	3.Enteric septicaemia of catfish	
4. Viral haemorrhagic septicaemia		
5. Epizootic ulcerative syndrome		
6. Red seabream iridoviral disease		
7. Infection with koi herpesvirus		
1.2 MOLLUSC DISEASES		
OIE-listed diseases	Non OIE-listed diseases	
1. Infection with Bonamia exitiosa	1. Infection with Marteilioides chungmuensis	
2. Infection with Perkinsus olseni	2. Akoya oyster disease	
3. Infection with abalone herpes-like virus	3. Acute viral necrosis (in scallops)	
4. Infection with Xenohaliotis californiensis		
1.3 CRUSTACEAN DISEASES		
OIE-listed diseases	Non OIE-listed diseases	
1. Taura syndrome	1. Monodon slow growth syndrome	
2. White spot disease	2. Milky haemolymph disease of spiny lobster	
3. Yellowhead disease	(Panulirus spp.)	
4. Infectious hypodermal and haematopoietic necrosis	3. Acute hepatopancreatic necrosis syndrome	
5. Infectious myonecrosis	(AHPNS)	
6. White tail disease (MrNV)		
7. Necrotising hepatopancreatitis		
1.4 AMPHIBIAN DISEASES		
OIE-listed diseases	Non OIE-listed diseases	
1. Infection with Ranavirus		
2. Infection with Bachtracochytrium dendrobatidis		
2. DISEASES PRESUMED EX	OTIC TO THE REGION	
2.1 Finfish		
OIE-listed diseases	Non OIE-listed diseases	
1. Infectious salmon anaemia	1. Channel catfish virus disease	
2. Infection with Gyrodactylus salaris		
2.2 Molluscs		
OIE-listed diseases	Non OIE-listed diseases	
1. Infection with Bonamia ostreae		
2. Infection with Marteilia refringens		
3. Infection with Perkinsus marinus		
2.3 Crustaceans		
OIE-listed diseases	Non OIE-listed diseases	
1. Crayfish plague (Aphanomyces astaci)		

Recent Aquatic Animal Health Related Publications

OIE Aquatic Animal Health Code, 15th Edition, 2012. The OIE Aquatic Animal Health Code (the Aquatic Code) sets out standards for the improvement of aquatic animal health and welfare and veterinary public health worldwide, including through standards for safe international trade in aquatic animals (amphibians, crustaceans, fish and molluscs) and their products. The health measures in the Aquatic Code should be used by the veterinary authorities of importing and exporting countries to provide for early detection, reporting and control of agents pathogenic to aquatic animals and, in the case of zoonotic diseases, for humans, and to prevent their transfer via international trade in aquatic animals and aquatic animal products, while avoiding unjustified sanitary barriers to trade. The health measures in the Aquatic Code have been formally adopted by the World Assembly of OIE Delegates, which constitutes the organisation's highest decision-making body. This 15th edition incorporates modifications to the Aquatic Code agreed at the 80th General Session in May 2012. The 2012 edition includes revised information on the following subjects: glossary; notification of diseases and epidemiological information; criteria for listing aquatic animal diseases; diseases listed by the OIE; import risk analysis; welfare of farmed fish during transport; welfare aspects of stunning and killing of farmed fish for human consumption; and disinfection of salmonid eggs for infectious haematopoietic necrosis, infectious salmon anaemia and viral haemorrhagic septicaemia. This edition includes four new chapters on communication; monitoring of the quantities and usage patterns of antimicrobial agents used in aquatic animals; development and harmonisation of national antimicrobial resistance surveillance and monitoring programmes for aquatic animals; and killing of farmed fish for disease control purposes. The Aquatic Animal Health Code is available for free download http://www.oie.int/en/international-standardsetting/aquatic-code/access-online/

OIE Manual of Diagnostic Tests for Aquatic Animals, 2013. The purpose of this manual is to provide a uniform approach to the detection of the diseases listed in the OIE *Aquatic Animal Health Code*, so that the requirements for health certification in connection with trade in aquatic animals and aquatic animal products can be met. It includes bibliographical references and a list of the OIE Reference Laboratories for amphibian, crustacean, fish and mollusc diseases. The manual is available for free download at http://www.oie.int/en/international-standard-setting/aquatic-manual/access-online/.

Tran, L., Nunan, L., Redman, R.M., Mohney, L.L., Pantoja, C.R., Fitzsimmons, K., Lightner, D.V., 2013. Determination of the infectious nature of the agent of acute hepatopancreatic necrosis syndrome affecting penaeid shrimp. Diseases of Aquatic Organisms, 105:45-55.

NACA, 2012. Final Report. Asia Pacific Regional Consultation on the Emerging Shrimp Disease – Early Mortality Syndrome (EMS)/Acute Hepatopancreatic Necrosis Syndrome (AHPNS). Network of Aquaculture Centres in Asia-Pacific, Bangkok, Thailand. <u>http://www.enaca.org/modules/library/publication.php?</u> publication_id=1059

OIE, 2012. Proceedings of OIE Global Conference on Aquatic Animal Health – Aquatic Animal Health Programmes: their Benefits for Global Food Security. World Organisation for Animal Health, Paris, France. 205 pp.

FAO, 2012. Improving biosecurity through prudent and responsible use of veterinary medicines in aquatic food production. FAO Fisheries and Aquaculture Technical Paper No. 547. FAO, Rome. 207 pp.

Leaño, E. M, and C.V. Mohan. 2012. Early mortality syndrome threatens Asia's shrimp farms. Global Aquaculture Advocate, July/August 2012: 38-39

Flegel, T.W., 2012. Historic emergence, impact and current status of shrimp pathogens in Asia. J. Invertebrate Pathology, 110:166-173.

Senapin, S., Phiwsaiya, K., Gangnonngiw, W., Flegel, T., 2011. False rumours of disease outbreaks caused by infectious myonecrosis virus (IMNV) in the whiteleg shrimp in Asia. Journal of Negative Results in BioMedicine, 10:10.

Rodgers, C.J., Mohan, C.V., Peeler, E.J., 2011. The spread of pathogens through trade in aquatic animals and their products. Rev. Sci. Tech, Off. Int. Epiz., 30: 241-256.

Jithendran, K.P., Shekar, M.S., Kannapan, S., Azad, I.S., 2011. Nodavirus infection in freshwater ornamental fishes in India: diagnostic histopathology and nested PCR. Asian Fisheries Science, 24:12-19.

Alday-Sanz, V., 2010. Chapter 24: **Designing a biosecurity plan at the facility level: criteria, steps and obstacles.** In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 655-678.

Benitez, J., Juarez, L., 2010. Chapter 30: The State Committees for Aquaculture Health: a success story from Mexico. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 821-833

Chen, S., Santos, M.D., Cowley, J., 2010. Chapter 28: What will PCR bring to shrimp farming: contribution, compromise or conflict. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 751-772.

Corsin, F., de Blas, N., 2010. Chapter 27: **Shrimp epidemiology: applying population-based methods to shrimp health management.** In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 713-749.

Cuellar-Anjel, J., Corteel, M., Galli, L., Alday-Sanz, V., Hasson, K.W., 2010. Chapter 22: **Principal shrimp infectious diseases, diagnosis and management**. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 517-621

Flegel, T.W., 2010. Chapter 23: Importance of host-viral interactions in the control of shrimp disease outbreaks. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 623-654.

Karunasagar, In., Karunasagar, Id., Alday-Sanz, V., 2010. Chapter 26: **Immunostimulants, probiotics and phage therapy: alternatives to antibiotics.** In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 695-711.

Lotz, J.M., 2010. Chapter 25: Evolutionary principles applied to disease control and health management in shrimp aquaculture. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 679-694.

Smith, P., 2010. Chapter 29: An economic framework for discussing antimicrobial agent use in shrimp farming. In: V. Alday-Sanz (ed), The Shrimp Book, Nottingham University Press. p. 773-820.

Lightner, D.V., Redman, R.M., 2010. The global status of significant infectious diseases of farmed shrimp. Asian Fisheries Science, 23:383-426.

Kono, T., Fall, J., Korenaga, H., Takayama, H., Iizasa, T., Mekata, T., Itami, T., Sakai, M., 2010. Immunomodulation by DNA vaccination against white spot syndrome virus (WSSV). Asian Fisheries Science, 23:435-446.

Sudhakaran, R., Mekata, T., Inada, M., Okugawa, S., Kono, T., Supamattaya, K., Yoshida, T., Sakai, M., Itami, T., 2010. Development of rapid, simple and sensitive real-time reverse transcriptase loop-mediated isothermal amplification method (RT-LAMP) to detect viral diseases (PRDV, YHV, IHHNV and TSV) of penaeid shrimp. Asian Fisheries Science, 23:561-575.

SEAFDEC AQD, 2010. **Prevention and Control of Parasites in Groupers** (Flyer). SEAFDEC Aquaculture Department, Tigbauan, Iloilo, Philippines. Available for free download at <u>http://www.seafdec.org.ph/</u>publications downloadable.html

Corsin, F., Georgiadis, M., Larry Hammel, K. and Hill, B., 2009. **Guide for Aquatic Animal Health Surveillance**. World Organization for Animal Health (OIE), Paris, France. 114 pp. Efficient and reliable surveillance systems generate sound evidence for disease incidence, prevalence and distribution, or for demonstrating disease absence. Science-based decisions regarding the health of aquatic animals rely on the information generated by surveillance programs. This practical handbook about surveillance is intended to be used mainly by Veterinary Services or other Competent Authorities, their staff and experts, for designing, implementing, and evaluating surveillance systems for diseases of relevance for aquatic animals in their country. The book can be ordered at http://www.oie.int/boutique/index.php?lang=en.

WHO-FAO Food Hygiene (Basic Texts), 4th Edition, 2009. World Health Organization and Food and Agriculture Organization of the United Nation, Rome, Italy. The Codex basic texts on food hygiene promote understanding of how rules and regulations on food hygiene are developed and applied. The General Principles of food hygiene cover hygiene practices from primary production through to final consumption, highlighting the key hygiene controls at each stage. This publication also contains the most internationally used description of the Hazard Analysis and Critical Control Point (HACCP) system and guidelines for its application. This fourth edition includes texts adopted by the Codex Alimentarius Commission up to 2009. The texts will be of use to government authorities, food industries, food handlers and consumers, as well as teachers and students of food hygiene.

Bondad-Reantaso, M.G., Arthur, J.R., Subasinghe, R.P. (eds), 2009. Strengthening Aquaculture Health Management in Bosnia and Herzegovina. FAO Fisheries and Aquaculture Technical Paper No. 524, Food an Agriculture Organization of the United Nation, Rome, Italy. 83 pp.

FAO, 2009. Report of the International Disease Investigation Task Force on a Serious Finfish Disease in Southern Africa. Food and Agriculture Organization of the United Nations, Rome, Italy. 70 pp.

FAO, 2009. What You Need to Know about Epizootic Ulcerative Syndrome: An Extension Brochure. Food and Agriculture Organization of the United Nations, Rome, Italy. 33 pp.

RECOFI. 2009. Proposal for a Regional Programme for Improving Aquatic Animal Health in RECOFI Member Countries. FAO Fisheries and Aquaculture Report No. 876, Food and Agriculture Organization of the United Nations, Rome, Italy. p. 101-118

Bondad-Reantaso, M.G., Arthur, J.R. and Subasinghe, R.P. (eds.). 2008. Understanding and applying risk analysis in aquaculture. *FAO Fisheries and Aquaculture Technical Paper*. *No. 519*. Rome, FAO. 2008. 304p. Risk analysis is an objective, systematic, standardized and defensible method of assessing the likelihood of negative consequences occurring due to a proposed action or activity and the likely magnitude of those consequences, or, simply put, it is "science-based decision-making"

FAO. Report of FAO **Workshop on Information Requirements for Maintaining Aquatic Animal Biosecurity.** Cebu City, Philippines, 15–17 February 2007. *FAO Fisheries and Aquaculture Report*. No. 877. Rome, FAO. 2008. 27p.

FAO Regional Commission for Fisheries. **Report of the Regional Technical Workshop on Aquatic Animal Health.** Jeddah. Kingdom of Saudi Arabia, 6-10 April 2008. FAO Fisheries and Aquaculture Report. No. 831. Rome, FAO. 2008. 120 pp.

FAO. 2009. Report of the International Emergency Disease Investigation Task Force on a Serious Finfish Disease in Southern Africa, 18-26 May 2007. Rome, FAO. 2009.

Arthur, J.R., Bondad-Reantaso, M.G. and Subasinghe, R.P. 2008. **Procedures for the quarantine of live aquatic animals: a manual**. FAO Fisheries Technical Paper No. 502. Rome, FAO. 2008. 74p.

Bondad-Reantaso, M.G., Mohan, C.V., Crumlish, M. and Subasinghe, R.P. (eds.) 2008. **Proceedings of the Sixth Symposium on Diseases in Asian Aquaculture (DAA VI)**. 25-28 October 2005, Colombo, Sri Lanka. Fish Health Section. 505 pp.

List of National Coordinators^{*}

Country	Name and Address	
Australia	Dr. Ingo Ernst Aquatic Animal Health Unit Office of the Chief Veterinary Officer Department of Agriculture, Fisheries and Forestry GPO Box 858, Canberra ACT 2601, Australia Fax: +61-2-6272 3150; Tel: +61-2-6272 4328 Email: <u>ingo.ernst@daff.gov.au</u> Dr. Herbert Brett Aquatic Animal Health Unit , Office of the Chief Veterinary Officer Department of Agriculture, Fisheries and Forestry GPO Box 858, Canberra ACT 2601, Australia Fax: +61 2 6272 3150; tel: +61 2 6272 4009	
Bangladesh	E-mail: brett.herbert@daff.gov.au Dr. M. G. Hussain Director General, Bangladesh Fisheries Research Institute (BFRI) Mymensingh 2201, Bangladesh Fax: +880-91-66559, Tel: +880-91-65874 E-mail: hussain.bfri@gmail.com; dg@fri.gov.bd; dgbfri@gmail.com	
Cambodia	Mr. Chheng Phen Acting Director Inland Fisheries Research and Development Institute (IFReDI) Fisheries Administration, # 186, Norodom Blvd., PO Box 582, Phnom Penh, Cambodia Phone: +855 23 221485 E-mail: chhengp@yahoo.com	
China	Mr. Zhuzewen Disease Prevention and Control Division National Fisheries Technique Extension Center (NFTEC) Ministry of Agriculture Mai Zi Dian Street No 18 Chanyang District, Beijing 100026, China Fax: +86-10-65074250; Tel: +86-10-64195073 E-mail:zewenzhu@sina.com	
DPR Korea	Mr. Chong Yong Ho Director of Fish Farming Technical Department, Bureau of Freshwater Culture Sochangdong Central District, P.O.Box. 95, Pyongyong, DPR Korea Fax: +850-2-814416; Tel: 3816001, 3816121	
Hong Kong China	Ms. Joanne On-on Lee Fisheries Officer (Aquaculture Management) 2 Agriculture, Fisheries and Conservation Department 8/F, Cheung Sha Wan Government Offices 303 Cheung Sha Wan Road, Kowloon, Hong Kong SAR Fax: +852 21520383; Tel: +852 21526808 E-mail: joanne oo lee@afcd.gov.hk	

^{*} The matrix provides a list of National Coordinators and focal points nominated by governments for the Asia-Pacific Quarterly Aquatic Animal Disease Reports.

India	Mr. Intisar Anees Siddiqui Fisheries Research & Investigation Officer Department of Animal Husbandry, Dairying and Fisheries Ministry of Agriculture, Krishi Bhawan, New Delhi 110114, India Tel: +91-11-23389419/23097013 Fax: +91-11-23070370/23384030 E-mail: intisarsiddiqui@yahoo.co.in		
Indonesia	Dr. Muhammad Murdjani Director, Fish Health and Environment Ministry of Marine Affairs and Fisheries Directorate General of Aquaculture JI. Harsono RM No. 3, Gedung Ps. Minggu Jakarta Selatan Indonesia 12550 Fax: +62-21-78835853; Tel: +62-21-7890552 E-mail: anna murdjani@yahoo.com.id		
Iran	Dr. Kazem Abdi Khazineh Jadid Director General, Aquatic Animal Health Department Iran Veterinary Organization Ministry of Jihad-E-Agriculture Seyed Jamaledin Asad-Abadi St., Vali-Asr Ave. P.O.Box 14155-6349, Tehran, Iran Tel: +98-21-88966877; Fax: +98-21-88957252 E-mail: kazemabdy@yahoo.com		
Japan	Mr. Toshiyuki Kubodera Director Fish and Fishery Products Safety Office Animal Products Safety Division Ministry of Agriculture, Forestry and Fisheries 1-2-1, Kasumigaseki Chiyoda-ku, Tokyo 100-8950, Japan Fax: +81 3 35012685; Tel: +81 3 67442105 E-mail: roshiyuki kubodera@nm.maff.go.jp		
Lao PDR	Mrs. Thongphoun Theungphachanh Quality Control Animal Product Department of Livestock and Fisheries DLF PO Box 811, Lao PDR Fax : +856 21 216380; Tel: +856 21 216380 or Mobile: +856 20 772 1115 Email: theungphachan@yahoo.com Dr. Bounthong Saphakdy Director of Fisheries Division Department of Livestock and Fisheries DLF P.O. Box 811, Lao PDR		
Malaysia	E-mail: <u>saphakdy@yahoo.com</u> Dr. Siti Zahrah Abdullah National Fish Health Research Centre 11960 Batu Maung Penang, Malaysia Fax: +60 4 6263977; Tel: +60 4 6263922 E-mail: <u>siti.zahrah.abd@gmail.com</u>		
Myanmar	Mr. U Saw Lah Pah Wah Department of Fisheries, Ministry of Livestock and Fisheries Sin Minn Road, Alone Township, Yangon, Myanmar Fax: +95 01 228-253; Tel: +95 01 283-304/705-547 E-mail: dof@mptmail.net.mm		

Nepal	Mr. Ram Prasad Panta	
	Senior Fisheries Development Officer	
	Central Fisheries Laboratory	
	Central Fisheries Building, Balaju, Machhapokhari	
	Kathmandu. Nepal.	
	Fax: +977 1 4350833; Tel: +977 1 4385854	
	E-mail: rppanta13@gmail.com	
Pakistan	Mr. Anser Mahmood Chatta	
	Deputy Fisheries Development Commissioner	
	Livestock Division, Ministry of Food, Agriculture and Livestock	
	10 th Floor, Shaheed-e-Millat Secretariat (Livestock Wing) I	
	Islamabad, Pakistan	
	Fax: +9251 9212630; Tel: +9251 9208267,	
	ansermchatta@yahoo.com	
Philippines	Dr. Joselito R. Somga	
	Aquaculturist II, Fish Health Section, BFAR	
	860 Arcadia Building, Quezon Avenue, Quezon City 1003	
	Fax: +63 2 3725055/4109987; Tel: +63 2 3723878 loc206 or 4109988 to 89	
	E-mail: jsomga@bfar.da.gov.ph	
Republic of	Dr. Myoung Ae Park	
Korea	Director, Pathology Division	
	National Fisheries Research and Development Institute	
	152-1, Haeanro, Gijang-up	
	Gijang-gun, Busan 619-705	
	Korea	
	Tel: +82-51-7202470	
	E-mail: mapark@nfrdi.go.kr	
Cinconoro		
Singapore	Mr. Hanif Loo Jang Jing	
	Programme Executive (Aquaculture)	
	Aquaculture Branch	
	Food Supply & Technology Department	
	Agri-Food & Veterinary Authority of Singapore	
	5 Maxwell Road, #01-00, Tower Block, MND Complex, Singapore 069110	
	Fax: +65 63257677; Tel: +65 63257636;	
	Email: loo jang jing@ava.gov.sg	
	Ms. Diana Chee	
	Aquatic Animal Health Branch	
	Animal and Plant Health Laboratories	
	6 Perahu Road, Singapore 718827	
	Fax: +65 63161090; Tel: +65 63165140	
	E-mail: Diana Chee@AVA.gov.sg	
Sri Lanka	Dr. Rajapaksa Arachilage Geetha Ramani	
	Veterinary Investigation Officer	
	Veterinary Investigation Center	
	Department of Animal Production and Health	
	Welisara, Sri Lanka	
	Tel: +94-112-9258213; +94-714-932169	
	E-mail: vic_welisara@yahoo.com	
Thailand	Dr. Jaree Polchana	
	E-mail: polchana@yahoo.com	
	Aquatic Animal Health Research Institute (AAHRI) Department of Fisheries , Kasetsart University Campus Jatujak, Bangkok 10900, Thailand Fax: +66 2 5613993; Tel: +66 2 5794122, 5796977 E-mail: <u>j polchana@yahoo.com</u>	

Vietnam	Dr. Le Van Khoa	
	Deputy Chief	
	Aquatic Animal Health Unit	
	Department of Animal Health (DAH)	
	15/78 Giai Phong Street, Dpng Da	
	Hanoi, Vietnam	
	Fax: +84 4 38685961; Tel: +84 4 38693605	
	E-mail: lvkhoa@dah.gov.vn	

Instructions on how to fill in the QUARTERLY AQUATIC ANIMAL DISEASE REPORT

(Revised during the Provisional Meeting of the AG¹, Bangkok, Thailand, November 7-9, 2001)

Symbols used in the report are similar to those used by FAO, OIE and WHO for the *Animal Health Yearbook*. Please read these instructions carefully before you fill in the forms.

Under the heading 'Country', please enter your country.

Under the heading 'Period', please enter the reporting quarter (months) and year, e.g. January to March 2002.

Under the heading "Month", please enter months of a quarter in question, e.g. January, February, March.

In "Level of Diagnosis", please enter the Level of Diagnosis used, e.g., I, II, or III. See Section C below.

In "Epidemiological Comment Numbers", please enter the serial numbers, and write your corresponding epidemiological comments on page 2. See Section D below for guidance on the subjects to be covered under Epidemiological Comments.

If an unknown disease of serious nature appears, please fill in the last line of the form, with additional information on "Level of Diagnosis" and "Epidemiological Comment Numbers" as above.

Please do not fail to enter "***" or "-" as appropriate against each disease, which is essential to incorporate your information on the *Quarterly Aquatic Animal Disease Report (Asia and Pacific Region.)*

If you have new aquatic animal health regulations introduced within the past six months, please describe them under Section 2 on page 2.

Please use the following symbols to fill in the forms.

A. Symbols used for negative occurrence are as follows:

*** This symbol means that no information on a disease in question is available due to reasons such as lack of surveillance systems or expertise.

- This symbol is used when a disease is not reported during a reporting period. However the disease is known to be present in the country (date of last outbreak is not always known).

0000 This symbol is used when disease surveillance is in place and a disease has never been reported.

(year) Year of last occurrence (a disease has been absent since then).

B. Symbols used for positive occurrence are shown below.

+ This symbol means that the disease in question is reported or known to be present.

+? This symbol is used when the presence of a disease is suspected but there is no recognised occurrence of clinical signs of the disease in the country. Serological evidence and isolation of the causal agent may indicate the presence of the disease, but no confirmed report is available. It is important that the species of animals to which it applies is indicated in the "Comments" on page 2 of the form if you use this symbol.

+() These symbols mean that a disease is present in a very limited zone or zones as exceptional cases. It may also include the occurrence of a disease in a quarantine area.

? This symbol is used only when a disease is suspected by the reporting officer, but the presence of the disease has not been confirmed.

+?() These symbols mean that confirmed infection/infestation is limited to one of more zones of the country, but no clinical disease.

?() These symbols mean the presence of the disease suspected but not confirmed in a zone.

¹ Regional Advisory Group on Aquatic Animal Health (AG)

C. Levels of Diagnosis

LEVEL	SITE	ACTIVITY
1	Field	Observation of animal and the environment Clinical examination
11	Laboratory	Parasitology Bacteriology Mycology Histopathology
111	Laboratory	Virology Electron microscopy Molecular biology Immunology

D. Subjects to be covered in the Epidemiological Comments

- 1. Origin of the disease or pathogen (history of the disease);
- 2. Mortality rate (high/low or decreasing/increasing);
- 3. Size of infected areas or names of infected areas;
- 4. Death toll (economic loss, etc.);
- 5. Preventive/control measures taken;
- 6. Disease characteristics (unusual clinical signs or lesions);
- 7. Pathogen (isolated/sero-typed);
- 8. Unknown diseases (describe details as much as possible);
- 9. Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); and
- 10. Published paper (articles in journals)/web site, etc.

IMPORTANT

Please send the **original report** or the best photocopy thereof to the OIE and/or NACA **by fax** and **registered airmail**. Faxed reports are needed to check whether or not the reports are all right. The deadline for submission of the reports is **two and a half months (75 days)** after the end of the quarterly period.

If you require further explanation, please write to the OIE (Tokyo), NACA (Bangkok) or FAO (Rome) at the following addresses, respectively:

OIE Regional Representation for Asia and the Pacific

Food Science Building 5F The University of Tokyo 1-1-1 Yayoi, Bunkyo-ku Tokyo, 113-8657, Japan Tel. +81 3 5805 1931; Fax +81 3 5805 1934 E-Mail: <u>rr.asiapacific@oie.int</u>

NACA

P. O. Box 1040, Kasetsart Post Office, Bangkok 10903, Thailand Tel: 66-2-561-1728/9 (ext. 117); Fax: 66-2-561-1727 Dr. E.M. Leaño E-mail: <u>eduardo@enaca.org</u>

FAO

Fishery Resources Division, Fisheries Department FAO of the United Nations Viale delle Terme di Caracalla, 00100 Rome Tel. +39 06 570 56473; Fax + 39 06 570 530 20 E-mail: <u>Rohana.Subasinghe@fao.org</u>

Notes

Published by the Network of Aquaculture Centres in Asia-Pacific and the Food and Agriculture Organization of the United Nations. For inquiries regarding editorial or technical content, please write to NACA, P.O. Box 1040, Kasetsart P.O., Bangkok 10903, Thailand; Tel. (662) 561-1728 to 9; Fax: (662) 561-1727; e-mail: info@enaca.org or eduardo@enaca.org. Website: http://www.enaca.org

ISSN 1513-6558